Surveillance of Drug-Resistant Tuberculosis in South-East Asia

Report of an Intercountry Training Course
Bangalore, India, 21-25 June 2004

WHO Project: ICP TUB 001

World Health Organization
Regional Office for South-East Asia
New Delhi
August 2004
## CONTENTS

<table>
<thead>
<tr>
<th>Page</th>
</tr>
</thead>
<tbody>
<tr>
<td>1. INTRODUCTION ........................................................................................................1</td>
</tr>
<tr>
<td>2. OBJECTIVES ........................................................................................................2</td>
</tr>
<tr>
<td>3. INAUGURAL SESSION ............................................................................................3</td>
</tr>
<tr>
<td>4. THE WORKSHOP .....................................................................................................3</td>
</tr>
<tr>
<td>4.1 Drug Resistance in Tuberculosis in the SEA Region ...........................................3</td>
</tr>
<tr>
<td>4.2 Sampling Techniques for Undertaking Surveillance ...........................................6</td>
</tr>
<tr>
<td>4.3 Laboratory Techniques for Resistance Determination with Emphasis on Quality Assurance ..................................................................................................................8</td>
</tr>
<tr>
<td>4.4 Mechanism for Networking and Sharing of Data with National Programme Managers .........................................................................................................................10</td>
</tr>
<tr>
<td>4.5 Country-specific Draft Plan of Action for Undertaking Surveillance of Drug Resistance in TB .........................................................................................................................10</td>
</tr>
<tr>
<td>5. RECOMMENDATIONS ...............................................................................................11</td>
</tr>
</tbody>
</table>

### Annexes

| 1. List of Participants ........................................................................................................12 |
| 2. Programme ...................................................................................................................15 |
1. INTRODUCTION

Tuberculosis (TB) is the biggest killer of young adults, the age at which they are economically most productive. Nine million new cases of TB occur every year. Two million people die annually of TB and that makes it the biggest killer after HIV. Almost 38% of the global TB burden is in the South-East Asia Region where annually 750,000 deaths occur due to TB, or 1500 everyday, or one every minute. Five of the 22 high burden countries are in the SEA Region. In 1993, WHO declared TB to be a global health emergency and a war against this scourge was initiated through the DOTS (Directly observed treatment, short course) strategy. DOTS and resistance in *Mycobacterium tuberculosis* have a relationship of cause and effect. Poorly implemented DOTS gives rise to greater resistance in tubercle bacilli and prevalence of resistant mycobacteria warrants management through DOTS plus, which is expensive, of longer duration and employs toxic drugs.

In spite of the success of DOTS, TB control programmes are faced with two major problems, HIV and drug resistance. Of the 6 million people living with HIV in the Region, 2.5 million are infected with TB. Nearly 60% of AIDS cases develop TB indicating that the latter is the most common life-threatening opportunistic infection associated with HIV. The occurrence of resistance in these cases will spell a certain and quick death for them. All the five high TB burden countries in the Region, with the exception of Bangladesh, are considered high burden for HIV too. This increases the incidence of co-infection of two major lethal infections.

One of the aims of ensuring effective management of tuberculosis is to minimize the development of drug resistance. Surveillance of anti-TB drug resistance is therefore an essential tool for monitoring the effectiveness of TB control programmes and improving national and global TB control efforts. Drug sensitivity tests for monitoring resistance have been developed and standardized.

In 1994, WHO joined forces with International Union Against Tuberculosis and Lung Diseases (IUATLD) and launched the Global Alliance on Antituberculosis Drug Resistance Surveillance. The aims of the global
The objectives of the training course were to:

1. Review the status of drug resistance in *Mycobacterium tuberculosis* in the countries of the SEA Region;
2. Discuss the techniques including sampling methodologies being followed in the Region in undertaking community-based drug resistance surveillance in tuberculosis;
3. Demonstrate resistance determination techniques with emphasis on quality assurance;
4. Formulate a mechanism for networking and sharing of data with programme managers, and
5. Develop a country-specific draft plan of action on drug surveillance.
3. **INAUGURAL SESSION**

The training course was inaugurated by Dr Prahlad Kumar, Director, National Tuberculosis Institute, Bangalore. He emphasized the importance of resistance in the efficient management of any tuberculosis control programme. Dr Kumar also described the role played by NTI in developing various strategies in TB control. Earlier the participants were welcomed by Dr V.H.Balasangameshwara, Head of the Mycobacteriology Laboratory of the Institute. Dr Rajesh Bhatia, Short-term Professional, SEARO, presented the objectives and expected outcome of the workshop. He also deliberated upon the role of WHO in advocating the strengthening of laboratories in drug resistance surveillance and promoting quality in these areas.

4. **THE WORKSHOP**

4.1 **Drug Resistance in Tuberculosis in the SEA Region**

Dr C.Paramasivan, Deputy Director, Tuberculosis Research Centre (TRC), Chennai gave an overview of the global problem of resistance in tuberculosis and shared the results of the third round of global disease surveillance. Drug resistance in previously not treated individuals (primary resistance) in three countries of the Region, viz Thailand, Nepal and India, ranged between 0.9% and 2.8%. The figures for India do not denote national figures since surveillance was undertaken in only three selected sites.

**Bangladesh**

The estimated new cases of smear-positive tuberculosis in Bangladesh are 105 per 100,000 population per year. A cure rate of 84% of the TB patients, against the target of 85%, and a case detection rate of 38% against the target of at least 70% of new smear-positive patients by the end of 2005 have been achieved. Bangladesh has adopted the WHO recommended DOTS strategy since 1993 to control TB and prevent MDR TB since 1993. It covers 98% of the total population. No national data are available for the prevalence of drug resistance tuberculosis at present. To increase CDR, intensified collaboration with the private sector, working places and prisons is necessary.
India

Drug resistance in new smear-positive pulmonary tuberculosis is common in India and varies widely from area to area. Data from TRC have shown a gradual increase in the prevalence of resistance to anti-tuberculosis drugs. For isoniazid, resistance has ranged from 3% to 17% and for streptomycin from 3% to 13%. Resistance to rifampicin started appearing in the 1990s and remains around 1%. Resistance to more than one drug has been observed and ranges from 0 to 7% for isoniazid and streptomycin <1% for other combinations. In a recent survey conducted in a rural district in Maharashtra, Wardha, the level of MDR TB among new cases was only 0.5%.

Drug resistance in previously treated pulmonary tuberculosis cases is higher than drug resistance in new cases. A study conducted by TRC in a district in Tamil Nadu, found an increase in the frequency of acquired drug resistance with 67% resistance to isoniazid, 26% to streptomycin and 12% to rifampicin and 6% to both isoniazid and rifampicin. Similar results have been reported from Delhi and Gujarat. Drug resistance in previously treated cases in Chennai was 63%, out of which 23.5% were resistant to single drug and 39.5% resistant to more than one drug.

Multi-drug resistance (MDR) in India is very low and ranges from 0 to 6%. In a recent report from Chennai on the prevalence of MDR-TB among patients undergoing treatment for varying periods of time in four TB sanatoria attached to four district tuberculosis centres in Tamil Nadu showed 20.3% to be harbouring multidrug-resistant strains. The majority of these patients had irregular and interrupted treatment owing to the non-availability of drugs.

Indonesia

Indonesia has implemented the DOTS strategy since 1995. A cure rate of more than 85% has been achieved. By the end 2003 the DOTS strategy has covered around 90% of existing health centres; however, most of the hospitals and the private health sector have not yet adopted the DOTS strategy. The case notification rate is around 40%, while it is estimated that at least 70% of the predicted new smear-positive TB cases are detected.

No countrywide data are currently available on resistance in TB. Some small-scale surveys in Jakarta have shown MDR levels of more than 4% in previously untreated cases, but it remains to be determined whether this
situation prevails throughout the country. A protocol on MDR surveillance following the guidelines for WHO’s global monitoring project has been developed and will be implemented in 2004-2005. In 2004 it is proposed to start drug resistance surveillance in East Java Province. This is expected to act as a surveillance model for other provinces.

Myanmar

During 2003, 77214 cases of TB were notified in the country with a notification rate of 154/100 000. The percentage of sputum smear-positive cases in new pulmonary TB cases is 51.3%. The estimated HIV-TB co infection was found to be 4.5% in a survey conducted during 1995-1997. MDR TB among new TB patients in an institution-based study during 1994-1995 was 1.25 %. In 2003, MDR TB among new TB patients was found to be 2.9%.

Nepal

Infection with TB is very common in Nepal. About 60% of the economically active adult population is infected with TB. The annual risk of tuberculosis infection (ARTI) is estimated to be 1.72%. It is also estimated that about 44 000 people develop tuberculosis annually. Nearly half of them (20 000) are sputum-positive for acid fast bacilli (AFB) and about 6,000-8,000 people die from tuberculosis every year. In 2002, 2.4% of TB patients were recorded to have HIV infection. Surveys on drug resistance show that 1.33% of new tuberculosis patients harbour MDR-TB. However, a steady decline has been observed in the rate of MDR-TB, from 3.7% in 1998 to 1.33 in 2001/2002. Surveillance from 1996 to 2001 showed that 41% of the pretreatment patients are resistant to at least one drug. Further, resistance increased from 9.8% in 1996-97 to 13.2% in 1998-99 but decreased to 11.0% in 2001-02. MDR-TB resistance was 1.2% in 1996-97, 3.7% in 1998-99 and 1.3% in 2001-02 among new cases.

Thailand

The National Tuberculosis programme (NTP) in Thailand was established in 1949. TB control was implemented as an integral part of the existing general health services as a permanent programme, aiming to provide countrywide service that is accessible to the population. The Directly Observed Treatment Short-Course (DOTS) was adopted in 1996, and countrywide DOTS coverage achieved in 2001.
In 2003, based on a population size of 63 million, the estimated total incidence for Thailand was reported as 137 per 100,000 population; sputum smear-positive TB cases numbered 38,000, the total number of TB cases was 86,000; the number of deaths from TB was 13,000, and the overall case notification rate was 60 per 100,000 population. Treatment success rate and retreatment rates were 77% and 4.95% respectively and the estimate of HIV-positive TB cases was 7.12%.

Drug resistance surveillance (DRS) was performed in 1997 and 2001. Randomly selected proportionate cluster sampling of patients was adopted countrywide. In 2001, DRS covered 66 clusters from 64 diagnostic centres in 55 of 75 provinces. About 1,677 cases from among new patients with previous treatment were diagnosed to be sputum smear-positive PTB, put on DOTS and cultures isolated from them were subjected to proportional method of susceptibility testing to four drugs, namely rifampicin, isoniazid, streptomycin and ethambutol.

The trend of drug resistance-tuberculosis has shown that MDR TB has decreased, from 2.02% in 1997 to 0.9% in 2003. Among the previously treated patients MDR-TB was shown in 20.3% cases.

4.2 Sampling Techniques for Undertaking Surveillance

Mr K.P. Unnikrishnan, Chief Statistician, NTI, described the importance of proper sampling for drug resistance surveillance, various techniques that are recommended by WHO and the basis of each of these. It was emphasized that the sampling universe for a survey on the prevalence of anti-TB drug resistance should include all newly-registered sputum smear-positive TB patients in the country. The calculation of an appropriate sample size should be based on the following:

- The total number of new sputum smear-positive cases detected in the previous year in the country or in the geographical setting to be studied;
- Expected prevalence of resistance to rifampicin from available data (in the absence of available data, the best guess of investigators should be used);
Precision should be between 1 and 2%, while ensuring that obtaining the calculated sample is logistically feasible;

A confidence interval of 95% should be used for estimated prevalence.

Different sampling strategies can be adopted to select a representative sample of TB patients. In order to select a representative group of newly registered patients, a randomization step is essential. Some of the most useful sampling strategies that were discussed were:

100% sampling of diagnostic centres
This sampling method is most suitable for small countries with relatively small numbers of TB diagnostic units, good infrastructure, and facilities for transportation of samples from all.

Cluster sampling
Cluster sampling methods are best used in situations where it is logistically difficult to cover the entire area of the country and where the number of TB diagnostic centres is high. With this design, centres are randomly selected, and all sputum smear-positive patients newly registered at these selected centres during a defined period of time are included in the survey. A defined intake period, which should not exceed 12 months, identical for all centres included in the survey, should result in a balanced sample, as centres are represented according to their burden of cases. This allows direct estimation of the prevalence of resistance from the proportion calculated in the sample. An unbiased estimate of the prevalence of drug resistance necessitates the inclusion of a specified minimum number of centres (i.e. 30) if there has to be a good probability of including centres of different types (clinics, dispensaries and hospitals of different sizes) scattered throughout the country.

Population-proportionate cluster sampling
To avoid the risk of drawing a sample that misses the largest diagnostic centres, a weighted cluster sampling technique can be used. Based on a list of all diagnostic centres with the numbers of newly-registered patients per year, a cumulative population list is compiled. Assuming that the minimum recommended number of 30 clusters is selected, the total number of patients registered per year in all the centres is divided by 30 to obtain the sampling interval.
The participants were provided with exercises for the sampling techniques, their problems discussed and sorted out.

4.3 **Laboratory Techniques for Resistance Determination with Emphasis on Quality Assurance**

Before the commencement of hands-on activities in the laboratory, the participants were briefed about the importance of quality, implementation of internal quality control measures, development of standard operating procedures, participation in external quality assessment schemes and integration of quality system in the culture of their respective laboratories to make them aware of the consequences of poor quality in any laboratory activity, especially the surveillance of drug-resistance in TB.

Presentation was also made by Dr Paramasivan on biosafety measures and shipment of infectious material that are essential in any laboratory that is undertaking work on Mycobacterium tuberculosis. The participants were taken around the newly-constructed BSL-III facility at the Institute to familiarize them with modern containment practices that are being considered as a prerequisite for handling drug resistance work on mycobacteria.

The participants were briefed about the availability of various techniques to determine the susceptibility of mycobacteria to antimicrobial agents. Drug susceptibility tests may be performed by either the **direct** or **indirect** method. For the **direct method**, the inoculum is a digested and decontaminated sputum (or other clinical specimen) in which AFB may be demonstrated in stained smears. In this method, the inoculum is truly representative of the bacillary population present in the specimen. The advantage of this method is that the drug susceptibility test results are available along with the culture results (by 3-4 weeks).

The **indirect test** is used for specimens that are smear-negative but culture-positive or when the growth in the control slope of the direct test is inadequate. Further, in the indirect test, the inoculum is standardized but at the same time is not truly representative and hence there is a chance of selecting a proportion of susceptible or resistant bacilli from the slope. For this reason, the inoculum is prepared by using a representative sweep of the entire surface of the growth on the slope.
The indirect sensitivity tests

There are three general methods used for determining drug susceptibility of mycobacteria by the indirect method: the absolute concentration method (MIC method), the resistance ratio (RR) method, and the proportion method. When properly standardized and performed, all three methods have been shown to be equally satisfactory.

The absolute concentration method uses a standardized inoculum grown on drug-free media and media containing graded concentrations of the drug(s) to be tested. Several concentrations of each drug are tested, and resistance is expressed in terms of the lowest concentration of the drug that inhibits growth; i.e., minimal inhibitory concentration (MIC). This method is greatly affected by the inoculum size and by the viability of the organisms.

The resistance ratio method compares the growth of unknown strains of tubercle bacilli with that of a standard laboratory strain (H$_{37}$Rv). Parallel sets of media, containing two-fold dilutions of the drug are inoculated with a standard inoculum prepared from both the unknown and standard strains of tubercle bacilli. Resistance is expressed as the ratio of MIC of the test strain to MIC for the standard strain in the same set. This test is also greatly affected by the inoculum size as well as the viability of the strains. In addition, any variation in the susceptibility of the standard strain also affects the resistance ratio of the test strain.

The proportion method enables a precise estimation of the proportion of mutants resistant to a given drug. Several 10-fold dilutions of inoculum are planted on to both control (drug-free) and drug-containing media; at least one dilution should yield isolated countable (50 -100) colonies. When these numbers are adjusted by multiplying by the dilution of inoculum used, the total number of viable colonies on the control medium, and the number of mutant colonies resistant to the drug concentrations tested may be estimated. The proportion of bacilli resistant to a given drug is then determined by expressing the resistant portion as a percentage of the total population tested. The proportion method is currently the method of choice and recommended by National Committee on Clinical Laboratory Standards (NCCLS), WHO and International Union Against Tuberculosis and Lung Diseases (IUATLD).
Various laboratory techniques for proportion method were first discussed in the lecture hall and subsequently demonstrated to the participants in the laboratory. These were:

- Pre-treatment of clinical material
- Preparation of Lowenstein Jensen (LJ) Medium
- Diluting various antimicrobial agents in recommended dilutions
- Inoculation onto LJ medium
- Reading the growth on LJ medium
- Calculating the ratio of resistant and sensitive strains
- Interpretation of results

Hands-on experience was also provided in important procedures and the problems experienced by participants discussed and resolved.

4.4 Mechanism for Networking and Sharing of Data with National Programme Managers

Dr Paramasivan described the importance of networking of laboratories and the structure and functions of existing network for drug resistance surveillance. Of the 22 supranational reference laboratories that exist in the world today, only one (Tuberculosis Research Centre, Chennai, India) is in the SEA Region. This supranational laboratory is linked with National Reference Laboratories for surveillance of drug resistance and also conducts external quality assessment for the national laboratories. It was suggested that the countries should have functional national reference laboratory which should forge strong linkages with the supranational reference laboratory as well as provincial/state laboratories. The use of modern tools of information technology should be accelerated to share data with national managers on a continuous basis to support the programme.

4.5 Country-specific Draft Plan of Action for Undertaking Surveillance of Drug Resistance in TB

Dr Rajesh Bhatia (WHO/SEARO) briefed the participants on the need for planning and the method of development of an action plan by identifying
various activities that are needed to achieve the specific objectives. Various parameters that need to be considered and included in the action plan are: description of the activity; time-frame, type of activity, person designated to undertake the same and the resources required to accomplish the activity, and supervision of the whole process. The participants developed country-specific draft action plans. Two of these were presented and discussed in the plenary session.

Various issues that need to be considered by the participants in the implementation of their respective plan of action were thoroughly discussed. Various technical and managerial problems raised by the participants were discussed and solutions suggested. Extensive discussions led to the formulation of recommendations that have been described in Section 5.

5. RECOMMENDATIONS

To Member Countries

(1) The knowledge and experience gained from the workshop in improving infrastructure and skills in the surveillance of drug resistance in TB should be utilized gainfully in supporting national efforts in TB control and strengthening of the DOTS programme.

(2) All laboratories should strive to implement an efficient quality system so that the results generated are reliable and match with the needs of the users.

(3) The Tuberculosis Research Centre, Chennai and the National Tuberculosis Institute, Bangalore, may be approached for any technical support and trouble-shooting in establishing surveillance of drug-resistance in TB.

To WHO

WHO should continue to provide technical support to all Member States in developing and strengthening infrastructure and skills in the surveillance of drug resistance in TB and should undertake independent appraisal of DRS in TB in the Region.
Annex 1

LIST OF PARTICIPANTS

Bangladesh

Dr Md Ziaul Karim
Medical Officer (BTC)
National Chest Diseases Institute and Hospital
Mohakhali
Dhaka
Phone: 0088-02-8856566, 0171-343663
Email: zia0107@dhaka.net

Dr Muhammad Mamunur Rashid
Senior Consultant
Chest Diseases Clinic
Sathkhira
Phone: 0088-0471-62300, 0171-310139
Email: rasheedm@bttb.net.bd

Dr Abdullah Al Mamun
Associate Professor (Medicine)
Khulna Medical College
Dhaka
Phone: 0088-0417-333330, 00880-171-385252
Email: ramamun@bttb.net.bd , aamamun296@yahoo.com

India

Professor CP Baveja
Department of Microbiology
Maulana Azad Medical College
New Delhi – 110 002
Phone: 011-27654150, 9811650452

Dr Ranjani Ramachandran
Senior Research Officer
Tuberculosis Research Centre
Chetput, Chennai 600 031
Phone: 044 – 28362432 Ext 258, 94440-57481
Email: rrwju15@hotmail.com

Dr Manpreet Bhalla
Senior Research Officer Microbiology
Lala Ram Swarup Institute of Chest Diseases
Sri Aurobindo Marg
New Delhi
Phone: 011-26854929 Ext - 155, 98996-90335

Indonesia

Dr Elly Trisnawati
Staff of Sub-Directorate Microbiology
Directorate-General of Medical Care
Ministry of Health
Jakarta
Phone: 6221-5221706, 28811811559 (Mobile)
Email: ellytris@yahoo.com

Dr Siti Nadia
National TB Programme
Medical Officer
Directorate-General of Communicable Disease Control & Environmental Health
Ministry of Health
Jakarta
Phone: 62-21-42877589 (O), 62-8128195313
Email: nadiawiweko@yahoo.com

Dr Maria M.P. SpPk
National TB Laboratory Officer
Directorate Laboratory Health
Ministry of Health
Jakarta
Phone: 52-31-5672192, 62-31-5053076
Email: tbcblksy@indosat.net.id
Surveillance of Drug-Resistant Tuberculosis in South-East Asia

Dr Sri Prihatini
WHO National Consultant
DRS Team
Surabaya
East Java
Phone: 62-31-8273193, 62-21-5204349,
62-818950085, 62-8123009141
Email: prihatinis@who.or.id,
sripb@yahoo.com

Myanmar
Dr Ti Ti
Sr Consultant Microbiologist
National TB reference
Laboratory National TB Programme
Yangon
Phone: 95-9-211914(R), 95-9-380953(O)
Email: ntp@baganmail.net.in

Dr Khin Htay Kyi
Consultant Microbiologist
National Health Laboratory
Mandalay
Phone: 95-9-380953(O)

Dr Tin Tin Mar
Assistant Microbiologist
Union Tuberculosis Institute
Yangon
Phone: 95-9-380953(O)

Nepal
Dr Naresh Pratap K.C.
Senior Medical Officer
Ministry of Health
Kathmandu
Phone: 977-1-4352164
Email: npkc@inos.com.np

Ms Sashi Kala Maskey
Lab Technician
Bhaktapur Hospital
Bhaktapur
Phone: 5523530

Mr Bholu Prasad Chaudhary
Lab Technician
National Tuberculosis Centre
Thimi, Bhaktapur
Phone: 6630706, 6630033(O)

Thailand
Mrs Dhanida Rienthong
Senior Medical Scientist
Bureau of AIDS, TB and STIs
Department of Disease Control
Nonthaburi
Thailand
Phone: 66-2-212-9187
Email: dhanida@health.moph.go.th

Mrs Sununtha Mongkoljit
Senior Medical Scientist Office of Disease Prevention & Control
Department of Disease Control
5, Nakhonratchasima Province
Nonthaburi
Thailand
Phone: 66-0-4421-2900
Email: sununtha3b@yahoo.com

WHO SEARO
Dr Rajesh Bhatia
STP-BCT
WHO, SEARO
New Delhi – 110 011
Email: Bhatiaraj@WHO SEA.ORG

Temporary Advisers
Dr C N Paramasivan
Deputy Director
Tuberculosis Research Centre
ICMR, Chetput
Chennai 600 031
Tele: 91-044-28362432
Email: sivamparam@yahoo.com
Local Organizer

Dr Prahlad Kumar
Director
National Tuberculosis Institute
Bellary Road
Bangalore
Tele: 91-080-23362431
Email: ntiindia@blr.vsnl.net.in

Facilitators

Dr. V H Balasangameshwara
Chief Medical Officer (NFSG) and
In-Charge Bacteriology Section
National Tuberculosis Institute
Bellary Road
Bangalore
Tele: 91-080-23441192/93/23447951, Ext: 113
Email: vhbs@vsnl.com, vhbs@yahoo.com

M r K P Unnikrishnan
Chief statistical Officer
National Tuberculosis Institute
Bellary Road
Bangalore
Tele: 91-080-23441192/93, Ext: 118
Email: kpuk_u@hotmail.com
## Annex 2

### Programme

**Monday - 21 June 2004**

<table>
<thead>
<tr>
<th>Time</th>
<th>Activity</th>
<th>Presenter(s)</th>
</tr>
</thead>
<tbody>
<tr>
<td>0830 - 0900 hrs</td>
<td>Registration &amp; Pre-test</td>
<td>Dr P Kumar, Director, NTI</td>
</tr>
<tr>
<td>0900 - 0930 hrs</td>
<td>Inauguration</td>
<td>Dr P Kumar, Dr Rajesh Bhatia, Dr Hans Kluge, WHO</td>
</tr>
<tr>
<td>0930 - 0945 hrs</td>
<td>Scope of Workshop</td>
<td>Dr Rajesh Bhatia</td>
</tr>
<tr>
<td>0945 - 1015 hrs</td>
<td>Establishing a drug resistance surveillance system</td>
<td>Dr P Kumar</td>
</tr>
<tr>
<td>1030 - 1230 hrs</td>
<td>Presentation of their country data</td>
<td>Participants (10 minutes for each country)</td>
</tr>
<tr>
<td>1230 - 1330 hrs</td>
<td>Drug resistance surveillance – Current scenario in SEAR Countries</td>
<td>Dr C N Paramasivan</td>
</tr>
<tr>
<td>1330 - 1500 hrs</td>
<td>Enhancing quality of Laboratory services</td>
<td>Dr C N Paramasivan</td>
</tr>
<tr>
<td>1500 - 1545 hrs</td>
<td>Lab Methods: Culture including pretreatment procedures</td>
<td>Dr C N Paramasivan</td>
</tr>
<tr>
<td>1600 - 1630 hrs</td>
<td>Preparation of Lowenstein-Jensen based drug media (Theory)</td>
<td>Dr V H Balasangameshwara</td>
</tr>
<tr>
<td>1630 - 1730 hrs</td>
<td>Preparation of Lowenstein-Jensen based drug media (Practical demonstration in Lab)</td>
<td>Dr V H Balasangameshwara/ Dr C N Paramasivan and NTI Lab staff</td>
</tr>
</tbody>
</table>

**Tuesday - 22 June 2004**

<table>
<thead>
<tr>
<th>Time</th>
<th>Activity</th>
<th>Presenter(s)</th>
</tr>
</thead>
<tbody>
<tr>
<td>0830 - 0930 hrs</td>
<td>Organization and survey outline</td>
<td>Dr V H Balasangameshwara</td>
</tr>
<tr>
<td>0930 - 1030 hrs</td>
<td>Sample size and sampling strategies</td>
<td>Mr K P Unnikrishnan</td>
</tr>
<tr>
<td>1045 - 1145 hrs</td>
<td>Exercises on Sample size and sampling strategies</td>
<td>Mr K P Unnikrishnan</td>
</tr>
<tr>
<td>1145 - 1330 hrs</td>
<td></td>
<td>Dr V H Balasangameshwara/ Dr C N Paramasivan</td>
</tr>
<tr>
<td>1430 - 1530 hrs</td>
<td><strong>Drug susceptibility testing Methods</strong></td>
<td>Dr C N Paramasivan</td>
</tr>
<tr>
<td>Time</td>
<td>Activity</td>
<td>Presenter(s)</td>
</tr>
<tr>
<td>--------------</td>
<td>---------------------------------------------------------------------------</td>
<td>------------------------------------------------------------------------------</td>
</tr>
<tr>
<td>1530 – 1730 hrs</td>
<td>Preparation of bacterial suspensions for proportion method: Practical demonstration</td>
<td>Dr V H Balasangameshwara/ Dr C N Paramasivan and NTI Lab staff</td>
</tr>
</tbody>
</table>

**Wednesday - 23 June 2004**

<table>
<thead>
<tr>
<th>Time</th>
<th>Activity</th>
<th>Presenter(s)</th>
</tr>
</thead>
<tbody>
<tr>
<td>0830 – 0900 hrs</td>
<td>Bio-safety Measures</td>
<td>Dr C N Paramasivan</td>
</tr>
<tr>
<td>0900 – 0930 hrs</td>
<td>Country presentation Bangladesh</td>
<td>BAN representative</td>
</tr>
<tr>
<td>0930 – 1100 hrs</td>
<td>EQA in smear microscopy:</td>
<td>Dr V H Balasangameshwara</td>
</tr>
<tr>
<td>1045 – 1145 hrs</td>
<td>Quality Control in culture and susceptibility testing of M. tuberculosis</td>
<td>Dr C N Paramasivan</td>
</tr>
<tr>
<td>1145 – 1330 hrs</td>
<td>Interpretation of results of proportion method: Theory</td>
<td>Dr V H Balasangameshwara</td>
</tr>
<tr>
<td>1430 – 1530 hrs</td>
<td>Interpretation of results of proportion method: Practical by participants</td>
<td>Dr V H Balasangameshwara/ Dr C N Paramasivan and NTI Lab staff</td>
</tr>
<tr>
<td>1545 – 1730 hrs</td>
<td>Drug Susceptibility testing methods for Non-tuberculosis mycobacteria</td>
<td>Dr C N Paramasivan</td>
</tr>
</tbody>
</table>

**Thursday - 24 June 2004**

<table>
<thead>
<tr>
<th>Time</th>
<th>Activity</th>
<th>Presenter(s)</th>
</tr>
</thead>
<tbody>
<tr>
<td>0830 – 0900 hrs</td>
<td>Safe shipment of specimens and cultures</td>
<td>Dr C N Paramasivan</td>
</tr>
<tr>
<td>0900 – 0930 hrs</td>
<td>Interpretation of Survey data results</td>
<td>Dr V H Balasangameshwara</td>
</tr>
<tr>
<td>0930 – 1000 hrs</td>
<td>Components of country specific DRS plan</td>
<td>Dr C N Paramasivan</td>
</tr>
<tr>
<td>1000 – 1030 hrs</td>
<td>How to develop a country Plan of Action</td>
<td>Dr Rajesh Bhatia</td>
</tr>
<tr>
<td>1045 – 1145 hrs</td>
<td>Development of country work plans</td>
<td>Groups</td>
</tr>
<tr>
<td>1145 – 1330 hrs</td>
<td>Identification tests for M. tuberculosis: Theory</td>
<td>Dr C N Paramasivan</td>
</tr>
<tr>
<td>1430 – 1700 hrs</td>
<td>Identification tests for M. tuberculosis: Practical demonstration</td>
<td>Dr V H Balasangameshwara/ Dr C N Paramasivan/ NTI Lab staff</td>
</tr>
<tr>
<td>1700 – 1730 hrs</td>
<td>Identification of need for repeat practical work for Day 5</td>
<td>Dr V H Balasangameshwara/ Dr C N Paramasivan</td>
</tr>
</tbody>
</table>
### Friday - 25 June 2004

<table>
<thead>
<tr>
<th>Time</th>
<th>Activity</th>
<th>Presenter(s)</th>
</tr>
</thead>
<tbody>
<tr>
<td>0830 - 0900 hrs</td>
<td>Post test</td>
<td></td>
</tr>
<tr>
<td>0900 - 1100 hrs</td>
<td>Suspension preparation and other lab work</td>
<td>Dr C N Paramasivan/ Dr V H Balasangameshwara</td>
</tr>
<tr>
<td>1100 - 1200 hrs</td>
<td>Presentation of country plan of actions</td>
<td>Dr Rajesh Bhatia</td>
</tr>
<tr>
<td>1200 - 1330 hrs</td>
<td>Panel discussion for trouble shooting</td>
<td>All faculty</td>
</tr>
<tr>
<td>1430 - 1630 hrs</td>
<td>Draft recommendations and valedictory</td>
<td>Director and All faculty</td>
</tr>
</tbody>
</table>