Chikungunya is a viral disease transmitted by Aedes mosquitoes. The disease typically consists of an acute illness characterised by fever, rash, and incapacitating arthralgia. The word chikungunya, which is used for both the virus and the disease, means “to walk bent over” in the African dialect Swahili or Makonde, and refers to the effect of the incapacitating arthralgia.4

Chikungunya is a specifically tropical disease. It is relatively uncommon and poorly documented.3,5 A chikungunya outbreak is currently ongoing in Réunion, in which Aedes albopictus is the presumed vector. In the ongoing Indian outbreak, in which Aedes aegypti is the presumed vector, 1,400,000 cases of chikungunya were reported during 2006. The reasons for the re-emergence of chikungunya on the Indian subcontinent, and for its unprecedented incidence rate in the Indian Ocean region, are unclear. Plausible explanations include increased tourism, chikungunya virus introduction into a naïve population, and viral mutation.

Introduction
Chikungunya is a viral disease transmitted by Aedes mosquitoes. The disease typically consists of an acute illness with fever, skin rash, and incapacitating arthralgia.1,2 The latter distinguishes chikungunya virus from dengue, which otherwise shares the same vectors, symptoms, and geographical distribution.2,3 The word chikungunya, which is used for both the virus and the disease, means “to walk bent over” in the African dialect Swahili or Makonde, and refers to the effect of the incapacitating arthralgia.4

Chikungunya is a specifically tropical disease. It is relatively uncommon and poorly documented.3,5 A chikungunya outbreak is currently ongoing in Réunion (Indian Ocean), where about 266,000 of the 775,000 inhabitants have reported symptoms of the disease. The probable vector is Aedes albopictus, a mosquito species endemic to Réunion and other islands in the Indian Ocean (figure 1A).1 In India, where the main vector is Aedes aegypti (figure 1B), 1,400,000 cases were reported during 2006.6 The last outbreak of the infection in India occurred in 1973.

Epidemiology and natural history Chikungunya virus
Chikungunya virus, an arbovirus belonging to the genus Alphavirus (Togaviridae family), has a single-stranded RNA genome, a 60–70 nm diameter capsid and a phospholipid envelope. It is sensitive to desiccation and to temperatures above 58°C.8,9 The Alphavirus group comprises 28 viruses, six of which can cause human joint disorders—namely chikungunya virus, o’nyong-nyong virus (central Africa), Ross River and Barmah Forest viruses (Australia and the Pacific), Sindbis virus (cosmopolitan), and Mayaro virus (South America, French Guiana).1 These alphaviruses share certain antigenic determinants.3,4 Three lineages with distinct genotypic and antigenic characteristics have been identified: isolates from the ongoing Indian Ocean outbreak represent a distinct clade within a large east central, and southern African phylogroup distinct from the Asian phylogroup and from the west African phylogroup.5 The current Indian epidemic is caused by the central/east African genotype. Chikungunya virus strains isolated in India are closely related to strains isolated in Réunion (99.61% nucleotide sequence homology) and to strains isolated in Maharashtra province in 2000 (98.95% homology).6 The viruses isolated in India from 1963 to 1973 belonged to the Asian genotype. All older isolates of the Asian genotype (India 1963–1973 and Thailand 1962–1978) cluster together, whereas later isolates from the Philippines (1985), Indonesia (1985), Thailand (1988, 1995–1996), and Malaysia (1988) form a distinct cluster.4

To date, ten complete chikungunya virus nucleotide sequences have been determined: two human isolates (Ross and S27) recovered during the 1952–1953 outbreak in Tanzania,8 one strain (#37997) originally isolated from Aedes furcifer during the Senegal outbreak in 1983,9 six during the recent outbreak in Réunion,10 and one (formally designated LR2006-OPY1) from a traveller returning from an Indian Ocean island.11 Schuffenecker and colleagues12 have reported the near-complete nucleotide sequences of chikungunya virus strains isolated from six patients in Réunion and the Seychelles, and partial (E1) sequences of isolates from 127 patients in Réunion, the Seychelles, Mayotte.
Madagascar, and Mauritius. Interestingly, these authors detected a mutation at residue 226 of the membrane fusion glycoprotein E1 (E1-A226 V), which was absent from the strains isolated during the first months of the ongoing outbreak in Réunion, but was found in more than 90% of isolates after September, 2005. This change could be related to virus adaptation to the mosquito vector species. Together with the lack of herd immunity, this might explain the abrupt and escalating nature of the Réunion outbreak. Other chikungunya isolates are currently being sequenced to provide spatiotemporal data on the viruses isolated from different locations (Réunion, Mayotte, Mauritius, Seychelles, and Madagascar) and to find out whether any molecular signatures could be associated with unusual clinical forms (Schuffenecker I, Centre National de Référence des Arbovirus, Institut Pasteur, Lyon, France, personal communication). A similar genetic change has been described in the Semliki Forest virus model.

The vast scale and sudden emergence of the Réunion outbreak underlines how little we know on the biology of chikungunya virus. It is also conceivable that chikungunya virus never disappeared entirely from the Indian subcontinent, and that the current outbreak is because of a simple resurgence.

Vectors
In Asia and the Indian Ocean region the main chikungunya virus vectors are A. aegypti and A. albopictus. A larger range of Aedes species (A. furcifer, Aedes vittatus, Aedes fulgens, Aedes luteocephalus, Aedes dalcizlei, Aedes vigilax, Aedes camptorhynchites) transmit the virus in Africa, and Culex annulirostris, Mansonia uniformis, and anopheles mosquitoes have also occasionally been incriminated.

A. albopictus has a wide geographical distribution, is particularly resilient, and can survive in both rural and urban environments. The mosquito’s eggs are highly resistant and can remain viable throughout the dry season, giving rise to larvae and adults the following rainy season. Originating from Asia, and initially sylvatic, A. albopictus has shown a remarkable capacity to adapt to human beings and to urbanisation, allowing it to supersede A. aegypti in many places (including China, the Seychelles, and Hawaii), and to become a secondary but important vector of dengue and other arboviruses. It seems that most new introductions of A. albopictus have been caused by vegetative eggs contained in timber and tyres exported from Asia throughout the world. A. albopictus is therefore both rural and urban, zoophilic and anthropophilic, thriving both in the natural environment, and in habitations and their immediate periphery. A. albopictus is relatively long-lived (4–8 weeks) and has a flight radius of 400–600 m. It is aggressive, silent, and diurnal, meaning that bednets are ineffective. The adult female appears to transmit the virus vertically to her eggs, although this remains to be documented in the Indian Ocean outbreak. A. albopictus has long been present in the Indian Ocean region, and was probably involved in the dengue epidemics that hit Réunion in 1977–78 and 2004. It is very difficult to avoid A. albopictus bites and to control this mosquito species in tropical, human-modified ecosystems with modern infrastructures, irrigation, and massive solid waste production. In India, the dominant carrier of chikungunya virus is A. aegypti, which breeds mainly in stored fresh water in urban and semi-urban environments.

Chikungunya virus is maintained in the human population by a human-mosquito-human transmission cycle that differs from the sylvatic transmission cycle described on the African continent. The cycle in Réunion probably follows a dengue-like model of outbreak dynamics, characterised by the absence of an animal reservoir and the ability to spread rapidly among human beings via domestic and peri-domestic mosquitoes.

Chikungunya virus has been imported to Europe and the USA by infected travellers returning from areas with high incidence rates, and A. albopictus has been introduced into several European countries (Belgium, Bosnia, Croatia, France, Greece, the Netherlands, Serbia, Spain, and Switzerland) and also to Central America, Brazil, and the USA. A. albopictus has been imported through the trade in used tyres and in ornamental plants. Some authors have suggested that if viraemic patients were to arrive in southern Europe during the summer they could cause a European outbreak.

Reservoirs
Human beings serve as the chikungunya virus reservoir during epidemic periods. Outside these periods the main reservoirs are monkeys, rodents, birds, and other unidentified vertebrates. Outbreaks might occur in monkeys when herd immunity is low; the animals develop viraemia but no pronounced physical manifestations.

Geographical range
Since the first recorded chikungunya epidemic, which occurred in Tanzania in 1952–53, human chikungunya virus infection has been documented in Burma, Thailand, Cambodia, Vietnam, India, Sri Lanka, and the Philippines. Epidemics were reported in the Philippines in 1954, 1956, and 1968, and in south Sumatra, Java, Timor, Sulawesi, and the Moluccas Islands between 1982 and 1985. 25 outbreaks were reported in Indonesia between 1999 and 2003, based on clinical observation (13 outbreaks) or serological diagnosis (12 outbreaks). Chikungunya occurs in west Africa, from Senegal to Cameroon, Democratic Republic of
the Congo, Nigeria, Angola, Uganda, Guinea, Malawi, Central African Republic, and Burundi, and also in southern Africa. Chikungunya is well known on the Indian subcontinent. Since the virus was first isolated in Calcutta in 1963 there have been several reports of chikungunya virus infection in India. Since December, 2005, it is estimated that more than 1400000 cases have occurred in India.

Neither Europe nor the Americas have had outbreaks of chikungunya virus so far. Imported cases have been diagnosed in the French West Indies and Guyana—where A aegypti is endemic—among travellers returning from Réunion (figure 2). Imported cases have also been reported in Germany, Switzerland, Italy, Norway, China, and French Guyana, and also among US Peace Corps volunteers in the Philippines.

Epidemics
As with all arboviruses, chikungunya virus outbreaks begin during the rainy season when vector density peaks. Available data suggest that chikungunya virus can be both endemic and epidemic. Schematically, the endemic form appears to affect mainly rural Africa, with a wide range of vector species and reservoirs, high-level and continuous transmission to largely immune populations, and small rural outbreaks or sporadic cases. In this situation, chikungunya virus may be discovered fortuitously during serological surveys of other arboviruses, since there is a high degree of cross-reactivity between tests for chikungunya virus and o’nyong-nyong virus.

By contrast, the epidemic form of chikungunya virus tends to be Asian and urban, and to be transmitted by two vectors (A aegypti and A albopictus) to populations with weak herd immunity. In these settings the disease is characterised by abrupt, massive epidemics with high attack rates (eg, 37% in India in 1978). The epidemics peak then decline gradually as an increasing proportion of the population develops immunity. This second scenario, which corresponds with the situation in Réunion, has previously been observed in Asia. In the 1960s, epidemics occurred in several distinct rural and urban foci, followed by a lull of 20–30 years and re-emergence in the 1990s in Thailand, Indonesia and the Philippines, and India. Bangkok, Jakarta, Rangoon, and Calcutta were greatly affected in the 1960s, with seroprevalence rates reaching 60–74% in adults. Economic development does not protect populations from vector-borne diseases such as dengue or chikungunya. On the contrary, development can favour outbreaks by profoundly modifying the ecosystem. Malaysia, a rich country by comparison with other countries affected by chikungunya, had its first recorded chikungunya virus epidemic in 1998. A aegypti is endemic in Malaysia, and the virus was probably introduced by migrant workers. The same vectors can sometimes transmit several arboviruses, and confusing mixed epidemics have occasionally been described, such as yellow fever plus chikungunya virus, dengue plus chikungunya virus, or more recently Plasmodium falciparum malaria plus dengue type 1 and chikungunya on Madagascar’s east coast.

Chikungunya epidemics probably occurred before the virus was discovered, as suggested by descriptions of epidemic fevers accompanied by pronounced arthralgia. They sometimes appear to have been confused with dengue, o’nyong-nyong, or Sindbis virus infection.

Recent outbreaks: India and Indian Ocean islands
The ongoing epidemic in the Indian Ocean region probably emerged first in Kenya (Lamu and Mombasa; July, 2004), before reaching the Comoros (January, 2005) and Seychelles (March, 2005), followed by Mauritius. Systematic studies showed a prevalence of 75% of the population in the Kenya (Lamu) outbreak, 63% in the Comoros, and 26% in Mayotte (2006). The virus reached Réunion (figure 2) in March–April, 2005, and around 266000 cases had been diagnosed by Feb 19, 2007. For the first time, a substantial number of
Indian chikungunya outbreak seems to have followed the outbreak in the Indian Ocean islands, and may be related to the heavy tourist traffic between the two regions. 

In a serosurvey done in Calcutta City 10 years ago, 4·37% of 379 samples tested positive for chikungunya virus. The highest rates were observed in the 51–55 year age-group, and no cases of seropositivity were detected in the youngest group.

Studies are underway to determine the genomic structure and virulence of Indian chikungunya virus isolates and to explain the re-emergence of chikungunya virus in this country.

The Indian and Indian Ocean outbreaks are caused by an African strain of chikungunya virus. The Kenyan outbreak in 2004–2006 followed several months of unusually warm and dry conditions along the Indian Ocean coastline, and such conditions may have favoured chikungunya virus transmission in the Indian Ocean islands.

**Chikungunya virus in travellers**

The Indian Ocean islands, India, and Malaysia are popular tourist destinations. According to the World Tourism Organization, an estimated 1 474 218 people travelled from Madagascar, Mauritius, Mayotte, Réunion, and the Seychelles to European countries in 2004. The outbreak in the Indian Ocean islands has substantially dented the region’s tourist industry.

Among travellers returning from the tropics, febrile arthralgia with exanthema may be caused by a variety of viral and bacterial infections, or adverse drug reactions.

Chikungunya and dengue are among the most difficult diseases to distinguish, especially because simultaneous co-infection can occur. In the only published study comparing the symptoms of chikungunya and dengue, done in Thailand, chikungunya symptom onset was more abrupt, fever was shorter lived, and rash, conjunctival injection, and arthralgia were more frequent than in dengue.

**Clinical manifestations**

**Common chikungunya virus infection**

After infection with chikungunya virus, there is a silent incubation period lasting 2–4 days on average (range 1–12 days). Clinical onset is abrupt (see table), with high fever, headache, back pain, myalgia, and arthralgia; the latter can be intense, affecting mainly the extremities (ankles, wrists, phalanges) but also the large joints. Skin involvement is present in about 40–50% of cases, and consists of (1) a pruriginous maculopapular rash predominating on the thorax, (2) facial oedema, or (3) in children, a bullous rash with pronounced sloughing, and (4) localised petechiae and gingivorrhagia (mainly in children). Haemorrhagic fever has been reported in chikungunya virus-infected patients in Thailand.

The table shows the frequency of signs and symptoms during the ongoing Réunion epidemic and the 1998

For more information on the chikungunya outbreak in Réunion see http://www.invs.sante.fr and http://www.onrsur.net

For more information on the European Centre for Disease Prevention and Control report on chikungunya see http://www.ecdc.eu.int

For more information on the chikungunya outbreak in India see http://mchfw.nic.in, http://who.int and http://www.hinduonnet.com
Malaysian epidemic. Symptoms generally resolve within 7–10 days, except for joint stiffness and pain,72,73

Arthralgia and arthritis

Erratic, relapsing, and incapacitating arthralgia is the hallmark of chikungunya, although it rarely affects children. It may persist for several months. There are few precise descriptions of chikungunya virus-associated joint disorders, and the underlying mechanism is unknown.17,72–74

More data are available on the arthritis caused by related alphaviruses, and especially Ross River virus infection, which has been described as an epidemic poly-arthritis.75,76,77,78 Arthralgia/arthritis appear to affect 73–80% of patients with serologically confirmed chikungunya virus or o’nyong nyong virus infection,72,74 and can persist in about 33% of patients for 4 months,76 15% for 20 months,73 and 10% for 3–5 years.72 Radiological findings are normal, and biological markers of inflammation are normal or moderately elevated.72,74

Severe forms affecting adults in Réunion

Chikungunya is not generally considered to be a life-threatening disease.39 As recently discussed by Stephen Higgs,23 by the summer of 2006 some 155 deaths might have occurred directly or indirectly caused by the disease in the ongoing Réunion outbreak. By March 14, 2006, the French health authorities had collated 96 serologically documented cases in Réunion residents who required intensive care; 79 of these patients were adults, and the male/female sex ratio was 1.4. 228 patients with a mean age of 78 years died.39 The possible link between chikungunya virus infection and multiorgan failure is under investigation.31,52

Among 3066 women that delivered children in Réunion hospitals during 2006, 159 reported symptoms of chikungunya virus infection had been notified in Réunion, of which 27 were severe.46 Neonatal chikungunya virus infection had never previously been reported. The possible risks of embryopathy, fetopathy, and late sequelae are unknown, and prospective follow-up of these “chikungunya virus babies” is therefore warranted. These cases are reminiscent of neonatal infection by dengue virus.46 Recently, Robillard and colleagues84 reported details of the first ten cases of maternofoetal chikungunya virus transmission and Lenglet and colleagues85 reported 16 cases of newborn babies presenting symptoms of, and confirmed diagnosis for, neonatal chikungunya.

Chikungunya in pregnancy and in newborn babies

Among 3066 women that delivered children in Réunion hospitals during 2006, 159 reported symptoms of chikungunya virus (124 during the pregnancy, 35 around the time of delivery), and chikungunya virus infection was confirmed by RT-PCR or serology in 151 (95%) of these women (data from Observatoire Régional de la Santé de La Réunion).

Among the 35 women who were ill at delivery, 30 delivered an infected newborn baby. As of March 16, 2006, 35 biologically documented cases of neonatal chikungunya virus infection had been notified in Réunion, of which 27 were severe.46 Neonatal chikungunya virus infection had never previously been reported. The possible risks of embryopathy, fetopathy, and late sequelae are unknown, and prospective follow-up of these “chikungunya virus babies” is therefore warranted. These cases are reminiscent of neonatal infection by dengue virus.46

Biological diagnosis of chikungunya virus infection

Virus isolation is based on inoculation of mosquito cell cultures, mosquitoes, mammalian cell cultures, or mice. Two main diagnostic methods are available, namely RT-PCR and serology (IgM or IgG) (figure 6).
RT-PCR is useful during the initial viraemic phase (day 0 to day 7), but classic serological methods are simpler (haemagglutination inhibition, complement binding, immunofluorescence, and ELISA). IgM is detectable after an average of 2 days by ELISA immunofluorescent assay (1–12 days) and persists for several weeks to 3 months. IgG is detected in convalescent samples and persists for years. The sensitivity and specificity of these tests are poorly established, and so is the possibility of false-positive reactions resulting from cross-reactivity with dengue or other arboviruses such as o’nyong-nyong virus. Serologically, chikungunya virus is most closely related to o’nyong-nyong virus and is a member of the Semliki Forest antigenic complex (panel).

Individual serological testing is not particularly useful, except when faced with atypical or severe forms, or in travellers returning from an epidemic zone. Parola and colleagues analysed sera from four infected travellers returning from Indian Ocean islands by using a quantitative real-time RT-PCR-based method. They detected a viral load of $10^9$ copies per mL in one case. Such high levels of viraemia are uncommon in arthropod-borne diseases such as dengue fever and West Nile disease.

PCR, antigen detection, and viral culture are also used to detect chikungunya virus in mosquitoes during epidemiological studies, and also to assess vector competence (the capacity to be infected by and transmit the virus).

Specific immunity and vaccination
Chikungunya virus infection seems to elicit long-lasting protective immunity. Experiments in animal models have shown cross-protection among chikungunya virus and other alphaviruses.

There is currently no commercial vaccine for chikungunya virus, although some candidate vaccines have been tested in human beings. In the latest trials, conducted by the US Army Medical Research Institute, very satisfactory seroconversion rates (98% on day 28) and neutralising antibody titres were obtained, persisting in 85% of cases at 1 year. Arthralgia occurred in 8% of the 59 volunteers. These American vaccine trials were interrupted in 2003, even though the candidate vaccine appeared safe in human beings, when other infectious disease research efforts were prioritised to counter potential terrorist use of biological agents. On Sept 6, 2006, the US Army Medical Research and Materiel Command signed a Material Transfer Agreement with the French National Institute of Health and Medical Research focusing on the records of previous clinical studies. A phase III trial of the US Army candidate vaccine is in preparation (US Embassy in France; press release, Sept 14, 2006). The candidate vaccine is a live...
vaccine (TSI-GSD-218) based on chikungunya virus strain 15561 isolated from a patient in Thailand and attenuated by serial passage in MRC-5 cells.

Prevention

 Pending vaccine development, the only effective preventive measures consist of individual protection against mosquito bites and vector control. Control of both adult and larval mosquito populations uses the same model as for dengue and has been relatively effective in many countries and settings. Mosquito control is the best available method for preventing chikungunya. Breeding sites must be removed, destroyed, frequently emptied, and cleaned or treated with insecticides. Large-scale prevention campaigns using DDT have been effective against A aegypti but not A albopictus. Control of A aegypti has rarely been achieved and never sustained. Recent data show the different degrees of insecticide resistance in A albopictus and A aegypti. However, vector control is an endless, costly, and labour-intensive measure and is not always well accepted by local populations, whose cooperation is crucial. Bednets should be used in hospitals and day-care facilities. Surveillance is also important for early identification of outbreaks. For example, in India, a multidisciplinary team was deployed in February, 2006, to assist local health authorities in improving public-health measures, including strengthening of arbovirus surveillance, clinical management of cases, vector control, and social mobilisation.

Treatment

There is currently no effective antiviral treatment for chikungunya. Treatment is therefore purely symptomatic and is based on non-salicylate analgesics and non-steroidal anti-inflammatory drugs. Synergistic efficacy was reported between interferon-α and ribavirin on chikungunya virus in vitro. A trial in southern Africa failed to confirm the efficacy of chloroquine on arthralgia.

Conclusion

Several lessons can be drawn from the ongoing outbreaks of chikungunya in India and the Indian Ocean islands. First, clinical manifestations are highly variable and may be more severe than previously reported. Second, economic development does not protect countries from vector-borne diseases (eg, West Nile virus in the USA, and dengue fever in Rio or Singapore); on the contrary, modern lifestyles may amplify an epidemic through travel, population ageing, and production of solid waste that can shelter aedes mosquitoes. Third, the chikungunya outbreaks highlight the importance of monitoring vector-borne and zoonotic diseases. These outbreaks have provided an opportunity to improve our previously poor knowledge of the biology, epidemiology, dynamics, and immunology of chikungunya virus infection. Genome sequencing may lead to new therapeutic and preventive measures.

Search strategy and selection criteria


Conflicts of interest

We declare that we have no conflicts of interest.

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Review


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